

Traditional Herbal Treatment for Plasmodium Falciparum: A Systematic Review on Traditional Plants to Treat Malaria in Iran

¹Ahmad Mehravaran, ²Hadi Mirahmadi, ³Leili Mohamadi, ⁴Hamid Dahmardeh

Abstract--

Background and Objective: Malaria is a vector-borne disease of global importance, with the vast majority of its life-threatening cases caused by infection with *Plasmodium falciparum* parasites. A marked decrease in malaria-related deaths worldwide has been attributed to the administration of effective antimalarial against *Plasmodium falciparum*. Considering the great potential of Iran in terms of plant biodiversity, rich traditional knowledge and practice, and malaria endemicity in the southeast of Iran. The present systematic review attempted to explore, synthesize and compile medicinal research findings on antimalarial plants in Iran.

Material and Method: The searches were conducted by three independent researchers to find the relevant studies published from January 1999 until the end of December 2018. We searched for published literature in the English language in MEDLINE via PubMed, EMBASE via Ovid, The Cochrane Library, and Trip database. The keywords used in the search strategy were malaria, *Plasmodium falciparum*, herbal medicine, traditional medicine, antimalaria treatment, anti-parasitic herbs.

Results: Detailed findings from five studies identified a total of Forty-one different plant species used in traditional malaria treatments throughout Iran. Khouzestan province represents the most antimalarial species, followed by Golestan. Aerial parts and flowers were the most frequently used plant parts. Six plants, including *Citrullus colocynthis*, *Buxushyrcana*, *physalis alkekengi*, *Glycyrrhiza glarba*, *Glycyrrhiza glabra*, and *Ferula oopoda* showed more antiplasmodial activity than others with IC50 values ranging from 2.01 to 26.6 µg/ml against K1 (Chloroquine-resistant) or 3D7 (Chloroquine-sensitive).

Conclusion: The investigated medicinal plants gathered in this article as a systematic review could be a key to identify the compounds with antimalarial effects.

Key words--Herbal treatment, Malaria, *Plasmodium Falciparum*, Plant biodiversity.

¹Infectious Disease and Tropical Medicine Research Center, Resistance Tuberculosis Institute, Zahedan University of Medical Sciences, Zahedan, Iran, Assistant Professor Department of Parasitology and Mycology, Faculty of Medicine, Zahedan University of Medical Sciences, Zahedan, Iran.

²Infectious Disease and Tropical Medicine Research Center, Resistance Tuberculosis Institute, Zahedan University of Medical Sciences, Zahedan, Iran, Assistant Professor Department of Parasitology and Mycology, Faculty of Medicine, Zahedan University of Medical Sciences, Zahedan, Iran

³PhD of Environmental Health, School of Medicine, Infectious Diseases and Tropical Medicine Research Center, Research Institute for Drug Resistant Tuberculosis, Zahedan University of Medical Sciences, Zahedan, Iran. Email: Lailimohamadi@gmail.com

⁴Assistant Professor of Radiology Zahedan University of Medical Sciences, Zahedan, Iran.

I. INTRODUCTION

Malaria is the most severe parasitic disease and one of the oldest recorded diseases in the world (1, 2). Malaria is the vast majority of its life-threatening cases caused by infection with *Plasmodium falciparum* parasites (3). It affects 219 million people per year worldwide (4). It is one of the world's leading health problems, causing about 435,000 deaths in 2017, the vast majority of deaths (99.7%) were due to *Plasmodium falciparum* malaria (5).

Malaria has now spread across a wide area around the equator, some regions of the Americas, Africa, and a large part of Asia. The importance of malaria in the World Economy and Health has led to important scientific projects that are considered to Evaluate and recognize the therapeutic solutions for this disease (6-9)

In recent years, the emergence of drug-resistant *Plasmodium* species has exacerbated the health and economic impact of malaria. In particular, *P. falciparum* (the most pathogenic human parasite) has developed resistance to virtually all currently available antimalarial drug (10). History shows that plants have been a pivotal source of medicines against malaria with two of the primary drugs used in malaria treatment, quinine and, more recently artemisinin, both having derived from traditional medicine and plants (11, 12). Artemisinin derivatives are now recommended by the World Health Organization worldwide. In combination with other drugs, such as lumefantrine, amodiaquine, mefloquine, sulphadoxine-pyrimethamine (SP), as the first-line treatment of malaria. This fact has encouraged the continuing search for new natural product-derived antimalarial drugs. There are several plants to treat malaria in endemic countries, also several studies have been conducted to evaluate the inhibitory effects of the plant extracts on *P.falciparum* (13-17). Traditional medicines are a potential supply source of new drugs against malaria and other infectious diseases (11). Using the medicinal products with plant origin has been extended due to fewer side effects, improving in patient acceptance, lower cost, and also more consistent with the normal physiological function of the human body (12). Despite the intensive efforts to control malaria, the disease continues to be one of the most significant health problems in the southeastern part of the country. Resistance to chloroquine for *P. falciparum* has been reported since 1983 in Iran.

Proper documentation of traditional medicine and plants used in the prophylaxis and treatment of malaria constitutes a critical task not only in preserving precious indigenous knowledge and biodiversity but also in enhancing community access to and stakes in the improvement of malaria control interventions. It is also crucial for stimulating future research on the safety and efficacy of medicinal plants and identification of chemical entities that could be developed into new standardized phytomedicines (18-30). Moreover, available research evidence on indigenous antimalarial plants is highly Scattered, which underscores the serious need for systematic compilation and synthesis. Considering the enormous potential of Iran in terms of plant biodiversity, rich traditional knowledge and practice, and malaria endemicity in the southeast of Iran, the present systematic review attempted to explore, synthesize and compile medicinal research findings on antimalarial plants in Iran.

II. MATERIAL AND METHOD

Search methods for eligible studies

Three independent researchers of the research team developed a strategy to search for articles to be included in the systematic review. Between 13th April and 18th June 2019 six medical databases, including Medline, Scopus, Excerpta Medica Database (EMBASE), Ovid, Trip database, and the Cochrane library, were used to search for potentially eligible publications. For literature published in Persian, we searched national databases (Magiran and SID). Supplemental sources included Boolean operators that helped to conduct more efficient searches from these databases. Also, search engines such as Google and Google Scholar were also used to identify all potentially eligible publications. Another search was performed between 7th February and 5th November 2019.

Eligibility criteria

The inclusion criteria we used to select articles are as follows: 1) Published medicinal surveys reporting on anti-Plasmodium falciparum plants conducted in Iran without time limitation; 2) Original retrospective and prospective blinded studies investigating the medicinal research findings on anti-Plasmodium falciparum plants in Iran; and 3) Published peer-reviewed and research articles between January 1999 and December 2018 and, written in English or Persian.

Exclusion criteria

The following types of research data were excluded from analysis: 1) Data from review articles, conference papers, preprints, historical documents or experimental studies; 2) Data from published and unpublished medicinal surveys lacking information on anyone of the following: study areas/localities, informant's involvement, scientific plant names, and not reporting information about antimalarial medicinal plants.

Search strategy

Three persons reviewed titles and abstracts of retrieved studies from medical database of interest. The full texts of each study were retrieved, analyzed, and the screened studies were included in the review. The special search strategies were created using the Health Sciences Librarian website with specialization in systematic review searches using the Medical Subject Headings (MeSH) phrases and open phrases in accordance with the PRESS standards. After finalizing the MEDLINE strategy, the results were compared to search the other databases. Similarly, PROSPERO was searched to find recent or ongoing systematic reviews. The key search terms, including malaria, *Plasmodium falciparum*, herbal medicine, traditional medicine, antimalaria treatment, and anti-parasitic herbs. Corresponding authors of relevant documents were asked to provide full texts when not free or inaccessible and when it was not possible, i.e., non-reply or negative reply from corresponding authors, these full texts were purchased. In addition, the reference list of relevant documents was also examined to increase the chances of finding eligible papers. The Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) flowchart was used to depict the entire stepwise process of the screening strategy.

Data extraction and risk of bias evaluation

The data were extracted for evaluating the characteristics of the participants and independently keyed in an Excel spreadsheet (Microsoft Office 2016, USA) by a person to ensure internal quality control of the database. The index test included characteristics, including specialized equipment, reference standard (executor of the tests, and the interval between tests). The information related to medicinal research findings on antimalarial plants in Iran was also extracted. The first reader extracted the data. The second and third readers confirmed the data, and he would have completed them if they were incomplete.

III. RESULTS

Study Selection

A total of 985 articles were extracted through our preliminary searches in different databases. Of the 465 non-redundant studies identified by analyzing the titles and abstracts, 432 studies were ruled out due to irrelevant titles. Of the existing 33 studies, 5 studies met the inclusion criteria, and of the 28 excluded studies, 18 did not have the full text, 5 were review articles, 2 were the letter to editor, and 3 did not meet the minimum inclusion criteria (Fig. 1).

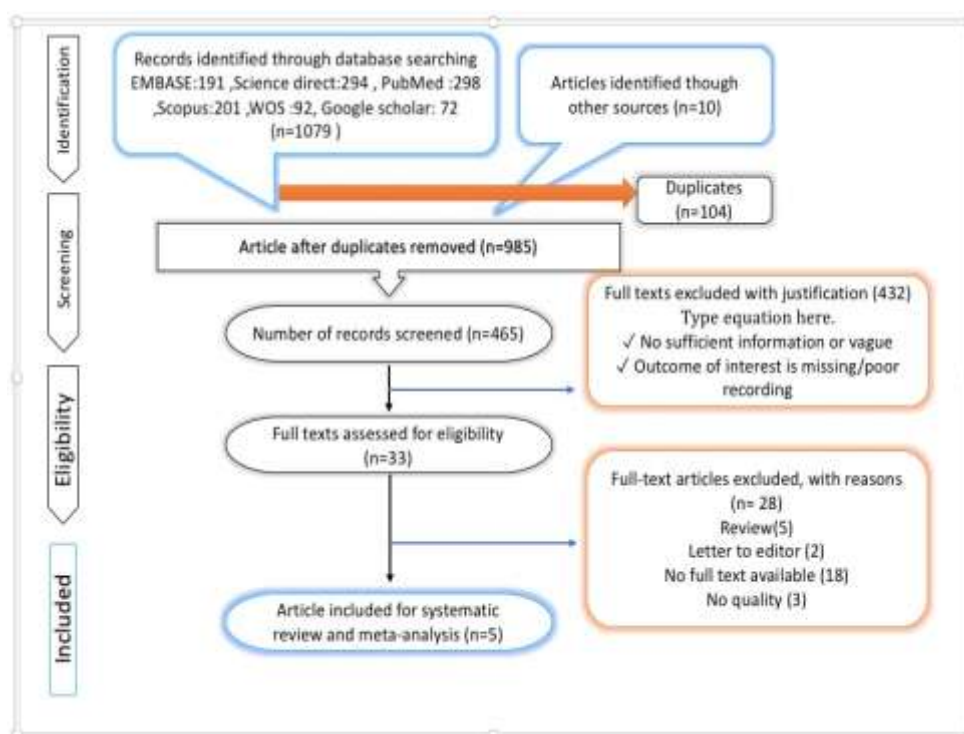


Fig 1. PRISMA flow diagram

Research Specifications

Forty-one different plant species used in traditional malaria treatments throughout Iran were studied. A total of 5 studies reporting the antimalarial species of 10 provinces meeting the inclusion criteria were reviewed. Of these 5 studies, three studies conducted in Tehran, Ahvaz and Zanjan. All of the studies were in vitro. Two of the studies were performed on the methanolic extracts of medicinal plants and the other three were done on the ethanolic extracts (Table 1).

Table 1. Included studies characteristics

ID	Author	Publication year	Province	Invitro/ Inivitro	Number of plant study	Extracted of
1	Esmaeili	2008	Tehran	Invitro	32	Methanol extracted
2	Ramazani	2010	Tehran	Invitro	10	Ethanol extracted
3	Ramazani	2010	Tehran	Invitro	5	Ethanol extractet
4	Sangian	2013	Zanjan	Invitro	10	Ethanol extracted
5	Hadaddad	2017	Ahvaz	Invitro	10	Methanol extracted

Anti- malarial medicinal plants in Iran

A total of 5 studies identified a total of Forty-one different plant species used in traditional malaria treatments throughout Iran. Table 1 summarizes the overall study characteristics. Table 2 summarizes the detail of traditional herbal medicine used for the treatment of malaria in Iran.



Fig 2. Geographic distribution of antimalarial plants in Iran

Geographic distribution of antimalarial plants

The geographic distribution of antimalarial plants is likely to be predicated on local trends concerning disease risk, floral diversity, and cultural diversity, including traditional medicinal practices. Khouzestan province represents the most antimalarial species, followed by Golestan (table 2).

Diversity of antimalarial plants

The antimalarial plant species identified in different region of Iran belonged to seventeen different plant families cited plant families included: Lamiaceae(16-18),Moraceae (16, 17), Solanaceae (16-18), Fabaceae (16, 17), Malvaceae (12, 18), Apiaceae (16), Asteraceae (16), Buxaceae (16), Cucurbitaceae (18), Salicaceae (18), Compositae (12, 18), Verbenaceae (18), Portulacaceae (18), Artemisia (19), Boraginaceae (12),

Papilionaceae (12), Plantaginaceae (12), Myrtaceae (12) Frequent citation of particular plant species or families could indicate potentially higher bioactive antimalarial content. Such evidence is pertinent for prioritizing future pharmacological research agendas. (table2)

Plant parts used and condition of preparations

The majority of antimalarial herbal remedies were prepared from a single plant part, while some were prepared from a combination of two or more plant parts. Aerial parts and flowers were the most frequently used plant parts (table2).

In vitro antiplasmodial activity

In the studies included, the anti-plasmodial activity of the extracts was determined against the chloroquine-resistant (K1) and chloroquine sensitive(3D7) strains of *Plasmodium falciparum* that were continuously cultured according to the methods described by Trager et al (20). Among these Forty-one plants, six plants showed better antiplasmodial activity: *Citrullus colocynthis* (cucurbitaceae), *Buxus hyrcana* (Buxaceae), *physalis alkekengi* (Solanaceae), *Glycyrrhiza glabra* (Papilionaceae), *Glycyrrhiza glabra* (Fabaceae) and *Ferula oopoda* (Apiaceae) with IC50 values ranging from 2.01 to 26.6 µg/ml against K1 or 3D7. (table 2)

Table 2. Medicinal plant families, species, locality, parts used, Date of collection, other medicinal values and Antiplasmodial activity (*Plasmodium falciparum*) of plants used for the treatment of malaria in Iran

Family	Scientific name	Locality (pro vince)	PU	Date of collection	Other medicinal values	Antiplasmodial activity <i>Plasmodium falciparum</i> , IC50 (µg/ml)		Referen ce
						Extracts IC50 3D7 (µg / ml)	Extracts IC50 K1 (µg / ml)	
Lamiaceae	<i>Otostegia persica</i> (Burm.) Boiss .	Sistan va Baluchestan	Fr, L	June 2008	Malaria, rheumatism	>64	31.1	16
	<i>Perovskia abrotanoides</i> Karel.	Golestan	AP	June 2008	Leshmaniasis, dermal disorders	>64	37.3	16
	<i>Eremostachys laciniata</i>	N/A	R	July 2010	Malaria	>200	N/A	18
	<i>Otostegia michauxii</i> Briq.	Fars	AP	June 2008	Malaria	>64	44.6	16
	<i>Teucrium polium</i>	Tehran	AP	July 2014	Fever, infection, inflammation	>200	>200	19
	<i>Marrubium vulgare</i> L	Mazandaran	B	N/a	Fever, bronchitis, vermifuge	>64	>64	16
	<i>Scutellaria multiscaulis</i> Boiss. subsp. <i>multicaulis</i>	Kohgiluyeh Buierahmad	AP	N/A	Antimicrobial	>64	>64	16
<i>Lavandula angustifolia</i>	Tehran	FL	August 2014	Fever, infection, inflammation	173.26	>200	19	

Solanaceae	<i>Solanum alatum</i>	Khouzestan	N/A	October, 2007	Headache, infection, wound healing, dysentery	>200	>200	17
	<i>Solanum surattense</i>	SistanvaBaluchestan	N/A	March, 2005	Antipyretic, inflammation	N/A	50	17
	<i>Solanum nigrum</i>	Guilan	Fr	September 2014	Fever, infection, inflammation	18.67	10.29	19
	<i>Physalis alkekengi</i>	Guilan	L,Fr	August 2014	Fever, infection, inflammation	13.08	11.31	19
Moraceae	<i>Ficus carica</i> L. var. <i>genuina</i> Boiss.	Tehran	AP	N/A	Hepatic tonic, malaria	>64	>54	16
	<i>Ficus bengalensis</i>	Khouzestan	N/A	May, 2007	Wound healing, sedative, antirheumatism, inflammation	>200	>200	17
	<i>Ficus carica</i>	Khouzestan	N/A	May, 2007	Ascites, Anemia, inflammation	>200	>200	17
	<i>Morus alba</i>	Khouzestan	N/A	May, 2007	Antipyretic, diuretic, laxative	>200	>200	17
Fabaceae	<i>Colutea persica</i> Boiss.	Mazandaran	AP	N/A	Antimicrobial	>64	>64	16
	<i>Glycyrrhiza glabra</i> L.	Gilan	AP	N/A	Gastric ulcers, hepatic disorders, malaria	>64	17.5	16
	<i>Prosopis juliflora</i>	Khouzestan	N/A	October, 2007	Fever, inflammation	N/A	14.78	17
	<i>Acacia farnesiana</i>	Khouzestan	N/A	May, 2007	wound healing, analgesic	>200	>200	17
Malvaceae	<i>Althea officinalis</i>	N/A	Fl	june 2010	Malaria	62.77	N/A	18
	<i>Gossypium herbaceum</i>	Golestan	L	July 2014	Fever, infection, inflammation	146.56	159.36	19
Apiaceae	<i>Asfrodacus orientalis</i> (L.) Drude.	Tehran	AP	N/A	Antimicrobial	42.6	46.1	16
	<i>Ferula oopoda</i> (Boiss and Bushe) Boiss.	Golestan	R	N/A	Cough, Migraine	24.9	26.6	16
Asteraceae	<i>Centaurea brugieriana</i> DC	Khorasan-e Shomali	AP	N/A	Antimicrobial	36.9	>64	16
	<i>Centaurea goletanica</i> (Akhani and Wagenitz.)	Golestan	AP	N/A	Antimicrobial	31.6	35.6	16
	<i>Matricaria chamomilla</i> L.	Tehran	AP	N/A	Dermal wound, inflammation, malaria	>64	>64	16
Buxaceae	<i>Buxus hyrcana</i> Poir.	Gilan	AP	N/A	Rheumatism, malaria	7.7	4.7	16

Cucurbitaceae	<i>Citrullus colocynthis</i>	Khouzestan	Fr	July 2014	Fever, infection, inflammation	2.01	6.9	19
Salicaceae	<i>Salix alba</i>	Alborz	L	July 2014	Fever, infection, inflammation	164.49	>200	19
Compositae	<i>Achillea millefolium</i>	Golestan	fl	May 2014	Fever, infection, inflammation	34.36	142.36	19
	<i>Anthemis nobilis</i>	N/A	fl	June 2012	Malaria	>200	N/A	18
	<i>Arctium lappa</i>	N/A	R	June 2010	Malaria	107.25	N/A	18
Verbenaceae	<i>Verbena officinalis</i>	Khozestan	Fl	June 2014	Fever, infection, inflammation	>200	>200	19
Portulacaceae	<i>Portulaca oleracea</i>	Tehran	AP	August 2014	Fever, infection, inflammation	136.57	172.39	19
Labiatae	<i>Melissa officinalis</i>	N/A	AP	June 2010	Malaria	>200	N/A	18
	<i>Stachys lavandulifolia</i>	N/A	Fl	June 2010	Malaria	156.49	N/A	18
Boraginaceae	<i>Borago officinalis</i>	N/A	Fl	May 2012	Malaria	62.77	N/A	18
Papilionaceae	<i>Glycyrrhiza glabra</i>	N/A	R	June 2012	Malaria	13.56	N/A	18
Plantaginaceae	<i>Plantago major</i>	N/A	S	May 2012	Malaria	40.00	N/A	18
Myrtaceae	<i>Myrtus communis</i>	N/A	AP	May 2012	Malaria	42.18	N/A	18

PU= Part Used (L=Leaf, S=Stem, R=Root, , AP= Aerial Part, Fr=Fruit, B:Bark, , S=Seed, Fl=Flower),N/A=not applicable

The resistance index is expressed as the ratio of the IC50 for strain K1 (Chloroquine resistant) to the IC50 for strain 3D7 Chloroquine-sensitive).

IV. DISCUSSION

This systematic review focused on studies having evaluated the antiplasmodial activity of plants in Iran. The findings of the present study showed that there are important medicinal plants against malaria in Iran that have the potential to inhibit malaria and are useful in removing *Plasmodium falciparum* infections. The results of this study are in line with those of other researchers around the world about the importance and understanding of antimalarial medicinal plant compounds (31-35).

Besides, the findings of our review study showed a high diversity of medicinal plants. This antimalarial plant species identified in different region of Iran belonged to Seventeen different plant families Cited plant families. Also, a large number of species have been observed in other studies in Africa and the United States (36, 37).

The antimalarial activity of herbal plants is due to the presence of a number of metabolically active compounds (37). These compounds include alkaloids (naphthyl isoquinolines, bisbenzyl isoquinolines, protoberberines and aporphines, indoles, manzamines, and miscellaneous alkaloids) terpenes (sesquiterpenes, triterpenes, diterpenes, and miscellaneous terpenes) quassinoids, flavonoids, limonoids, chalcones, peptides, xanthenes, quinones and coumarines, and miscellaneous antimalarials from nature (38-44).

The solvent of extraction largely determines the concentrations of the active metabolites in the extract. For example, methanolic extracts of the herbal plants are more active in vitro than water extracts, probably due to the presence of higher amounts of more active lipophilic compounds. The levels of activity of the antimalarial plant extracts depend on the concentration of the active antimalarial secondary metabolites. For example, gedunin, a very active compound against Plasmodium present in leaves of *A. indica* showed an IC₅₀ of 0.02 µg/ml against *P. falciparum*, but its concentration in the plant is very low and thus moderate activity of its extract (32, 45, 46).

The manner and conditions of the use of antimalarial medicinal plants are essential. Effective constituents of medicinal plants are reduced under inappropriate conditions, and proper use of medicinal herbs is vital for disease recovery.

Most commonly, the herbal medicines are prepared as water extracts in the form of decoction and infusion or as steam baths. The dose of the extract given is dependent on the age of the patient and the “strength” of the herbal medicine, although occasionally the weight of the patient. The quantity of extract given ranges from 100 to 500 ml, 100 to 250 ml, and 1 to 3 tea or tablespoons for adults, older children, and young children below five years of age, respectively, between 1 and 3 times a day for about a week or until when the patient has recovered. The extracts are mostly prepared from single herbal plants or from the combination of two herbal plants, for example, a decoction of *Tamarindus indica* and *Mangifera indica* is common. Medicine for malaria treatment from a herb such as *B. pilosa* can be made by squeezing a handful of its freshly picked leaves and drinking 1–3 teaspoons of the extract a day. Occasionally, malaria herbal medicines can be obtained by preparing different plant parts in combination; for example, an infusion can be made from fresh leaves and pounded fresh roots of *V. amygdalina* (32, 46-50).

Now, the important problem with malaria is the mutations in the back of the head that cause the resistance of some *Plasmodium falciparum* strains to many drugs, including artemisinin (50-53). Genetic studies have shown that *Plasmodium falciparum* isolated from the Amazon forest possesses mutant drug resistance genes that affect treatment (54).

In this studies, the antiplasmodial activity of the extracts was determined against the chloroquine-resistant and chloroquine-sensitive strains of *Plasmodium falciparum*. Six plants including: *Citrullus colocynthis*, *Buxushyrcana*, *physalis alkekengi*, *Glycyrrhiza glarba*, *Glycyrrhiza glabra* and *Ferula oopoda* showed more antiplasmodial activity with IC₅₀ values ranging from 2.01 to 26.6 µg/ml against K1 or 3D7. Similar studies have investigated the antiplasmodial activity of medicinal plants and obtained similar results (55-62).

The suggestion of this study is to introduce a new drug for the control of malaria using medical and herbal sciences and combine the different herbal species in Iran and other parts of the world. Further studies and

in vivo studies on the efficacy of these herbal remedies in the in vivo environment are needed to determine the effective concentration of these herbal remedies on clinical strains, their side effects, and their optimal performance. Evaluation to finally introduce these strains as a new anti-Plasmodium drug after completion.

V. CONCLUSION

It provides an overview of the antiplasmodial potential of medicinal Iran plants and highlights their usefulness as promising sources for new antimalarial drugs. The investigated medicinal plants gathered in this article as a systematic review could be a key to identify the compounds with antimalarial effects; therefore, if their compounds are examined, they might help to develop new and more efficient drugs.

It is hoped that such research can be used effectively in different parts of society to achieve better and healthier results and that the results of theoretical and laboratory studies will be applied to the world of medicine and industry.

Declaration of interest

The authors report no conflicts of interest.

Funding

This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors

REFERENCES

1. Kalra BS, Chawla S, Gupta P, Valecha N. Screening of antimalarial drugs: An overview. *Indian journal of pharmacology*. 2006;38(1):5.
2. Mirahmadi H, Fallahi S, Tabaei SJS. Soluble recombinant merozoite surface antigen-142 kDa of *Plasmodium vivax*: an improved diagnostic antigen for vivax malaria. *Journal of microbiological methods*. 2016;123:44-50.
3. Keeling P, Rayner J. The origins of malaria: there are more things in heaven and earth.... *Parasitology*. 2015;142(S1):S16-S25.
4. Organization WH. Malaria: fact sheet. World Health Organization. Regional Office for the Eastern Mediterranean; 2014.
5. Weiss DJ, Lucas TC, Nguyen M, Nandi AK, Bisanzio D, Battle KE, et al. Mapping the global prevalence, incidence, and mortality of *Plasmodium falciparum*, 2000–17: a spatial and temporal modelling study. *The Lancet*. 2019;394(10195):322-31.
6. Hay SI, Okiro EA, Gething PW, Patil AP, Tatem AJ, Guerra CA, et al. Estimating the global clinical burden of *Plasmodium falciparum* malaria in 2007. *PLoS Med*. 2010;7(6):e1000290.
7. Nadjm B, Behrens RH. Malaria: An update for physicians. *Infectious Disease Clinics*. 2012;26(2):243-59.
8. Caraballo H, King K. Emergency department management of mosquito-borne illness: malaria, dengue, and West Nile virus. *Emergency medicine practice*. 2014;16(5):1-23; quiz -4.
9. Brabin B, Dorman E, Beales P. Severe falciparum malaria. World Health Organization, Communicable Diseases Cluster. *Transactions of the Royal Society of Tropical Medicine and Hygiene*. 2000.
10. Malaria W. Fact sheet, Geneva: World Health Organization; 2018. 2019.
11. Ebrahimzadeh A, Dalir SN, Mirahmadi H, Mehravaran A, Khorashad AS, Turki H. The incidence of current infection with different human malaria species by polymerase chain reaction for diagnosis of suspicious malaria patients on elimination region Sistan and Baluchistan province, southeast of Iran. *Jundishapur Journal of Microbiology*. 2017;10(10).

12. Ashley EA, Dhorda M, Fairhurst RM, Amaratunga C, Lim P, Suon S, et al. Spread of artemisinin resistance in *Plasmodium falciparum* malaria. *New England Journal of Medicine*. 2014;371(5):411-23.
13. Malaria RB, Organization WH. The use of antimalarial drugs. Report of a WHO informal consultation Geneva: WHO. 2001.
14. Mutabingwa TK. Artemisinin-based combination therapies (ACTs): best hope for malaria treatment but inaccessible to the needy! *Acta tropica*. 2005;95(3):305-15.
15. Le Tran Q, Tezuka Y, Ueda J-y, Nguyen NT, Maruyama Y, Begum K, et al. In vitro antiplasmodial activity of antimalarial medicinal plants used in Vietnamese traditional medicine. *Journal of Ethnopharmacology*. 2003;86(2-3):249-52.
16. Wanyoike G, Chhabra S, Lang'at-Thoruwa C, Omar S. Brine shrimp toxicity and antiplasmodial activity of five Kenyan medicinal plants. *Journal of Ethnopharmacology*. 2004;90(1):129-33.
17. Abai M, Mehravaran A, Vatandoost H, Oshaghi M, Javadian E, Mashayekhi M, et al. Comparative performance of imagicides on *Anopheles stephensi*, main malaria vector in a malarious area, southern Iran. *J Vector Borne Dis*. 2008;45(4):307-12.
18. Takala-Harrison S, Jacob CG, Arze C, Cummings MP, Silva JC, Dondorp AM, et al. Independent emergence of artemisinin resistance mutations among *Plasmodium falciparum* in Southeast Asia. *The Journal of infectious diseases*. 2015;211(5):670-9.
19. Tun KM, Imwong M, Lwin KM, Win AA, Hlaing TM, Hlaing T, et al. Spread of artemisinin-resistant *Plasmodium falciparum* in Myanmar: a cross-sectional survey of the K13 molecular marker. *The Lancet infectious diseases*. 2015;15(4):415-21.
20. Wang J, Zhang C-J, Chia WN, Loh CC, Li Z, Lee YM, et al. Haem-activated promiscuous targeting of artemisinin in *Plasmodium falciparum*. *Nature communications*. 2015;6(1):1-11.
21. Ghorbal M, Gorman M, Macpherson CR, Martins RM, Scherf A, Lopez-Rubio J-J. Genome editing in the human malaria parasite *Plasmodium falciparum* using the CRISPR-Cas9 system. *Nature biotechnology*. 2014;32(8):819.
22. Mok S, Ashley EA, Ferreira PE, Zhu L, Lin Z, Yeo T, et al. Population transcriptomics of human malaria parasites reveals the mechanism of artemisinin resistance. *Science*. 2015;347(6220):431-5.
23. Ataide R, Ashley EA, Powell R, Chan J-A, Malloy MJ, O'Flaherty K, et al. Host immunity to *Plasmodium falciparum* and the assessment of emerging artemisinin resistance in a multinational cohort. *Proceedings of the National Academy of Sciences*. 2017;114(13):3515-20.
24. Veiga MI, Dhingra SK, Henrich PP, Straimer J, Gnädig N, Uhlemann A-C, et al. Globally prevalent PfMDR1 mutations modulate *Plasmodium falciparum* susceptibility to artemisinin-based combination therapies. *Nature communications*. 2016;7(1):1-12.
25. Ménard D, Khim N, Beghain J, Adegnik AA, Shafiul-Alam M, Amodu O, et al. A worldwide map of *Plasmodium falciparum* K13-propeller polymorphisms. *New England Journal of Medicine*. 2016;374(25):2453-64.
26. Straimer J, Gnädig N, Witkowski B, Amaratunga C, Duru V, Ramadani A, et al. Drug resistance. K13-propeller mutations confer artemisinin resistance in. 2015.
27. Karamati SA, Hassanzadazar H, Bahmani M, Rafieian Kopaei M. Herbal and chemical drugs effective on malaria. *Asian Pacific Journal of Tropical Disease*. 2014;4(S2):S599-S601.
28. Edrissian G, Shahabi S, Pishva E, Hajseyed-Javadi J, Khaleghian B, Ghorbani M, et al. Imported cases of chloroquine-resistant falciparum malaria in Iran. *Bulletin de la Societe de pathologie exotique et de ses filiales*. 1986;79(2):217-21.
29. Hunde D, Asfaw Z, Kelbessa E. Use of traditional medicinal plants by people of 'Boosat' sub district, Central Eastern Ethiopia. *Ethiopian Journal of Health Sciences*. 2006;16(2).
30. Esmaeili S, Naghibi F, Mosaddegh M, Sahranavard S, Ghafari S, Abdullah NR. Screening of antiplasmodial properties among some traditionally used Iranian plants. *Journal of Ethnopharmacology*. 2009;121(3):400-4.
31. Abebe E. Ethnobotanical study on medicinal plants used by local communities in Debark Wereda, North Gondar Zone, Amhara Regional State, Ethiopia: Addis Ababa University; 2011.
32. Philip K, Elizabeth MM, Cheplogoi PK, Samuel KT. Ethnobotanical survey of antimalarial medicinal plants used in Butebo County, Eastern Uganda. *European Journal of Medicinal Plants*. 2017:1-22.
33. Azas N, Laurencin N, Delmas F, Di Giorgio C, Gasquet M, Laget M, et al. Synergistic in vitro antimalarial activity of plant extracts used as traditional herbal remedies in Mali. *Parasitology research*. 2002;88(2):165-71.
34. Obbo C, Kariuki S, Gathirwa J, Olaho-Mukani W, Cheplogoi P, Mwangi E. In vitro antiplasmodial, antitrypanosomal and antileishmanial activities of selected medicinal plants from Ugandan flora: refocusing into multi-component potentials. *Journal of ethnopharmacology*. 2019;229:127-36.

35. Muganza DM, Fruth B, Lami JN, Mesia G, Kambu O, Tona G, et al. In vitro antiprotozoal and cytotoxic activity of 33 ethnopharmacologically selected medicinal plants from Democratic Republic of Congo. *Journal of ethnopharmacology*. 2012;141(1):301-8.
36. Silva JRdA, Ramos AdS, Machado M, de Moura DF, Neto Z, Canto-Cavalheiro MM, et al. A review of antimalarial plants used in traditional medicine in communities in Portuguese-speaking countries: Brazil, Mozambique, Cape Verde, Guinea-Bissau, São Tomé and Príncipe and Angola. *Memorias do Instituto Oswaldo Cruz*. 2011;106:142-58.
37. Alebie G, Urga B, Worku A. Systematic review on traditional medicinal plants used for the treatment of malaria in Ethiopia: trends and perspectives. *Malaria journal*. 2017;16(1):307.
38. Waako P, Katuura E, Smith P, Folb P. East African medicinal plants as a source of lead compounds for the development of new antimalarial drugs. *African Journal of Ecology*. 2007;45:102-6.
39. Chierrito TPC, Cunha A, De C, Koike L, Gonçalves R, Oliveira A, et al. Use of associated chromatographic techniques in bio-monitored isolation of bioactive monoterpene indole alkaloids from *Aspidosperma ramiflorum*. *Chromatography and Its Applications*. 2012;7:119-30.
40. Bhatnagar S, Das P. Antimalarial activity in tropical plants: a review. *Journal of herbs, spices & medicinal plants*. 2007;13(1):103-32.
41. Barku V, Opoku-Boahen Y, Dzotsi E. Isolation and pharmacological activities of alkaloids from *Cryptolepis sanguinolenta* (Lindl) Schl. *Int Res J Biochem Bioinform*. 2012;2:58-61.
42. e Silva LR, Montoia A, Amorim R, Melo M, Henrique M, Nunomura SM, et al. Comparative in vitro and in vivo antimalarial activity of the indole alkaloids ellipticine, olivacine, cryptolepine and a synthetic cryptolepine analog. *Phytomedicine*. 2012;20(1):71-6.
43. dos Santos Torres ZE, Silveira ER, Rocha e Silva LF, Lima ES, De Vasconcellos MC, de Andrade Uchoa DE, et al. Chemical composition of *Aspidosperma ulei* Markgr. and antiplasmodial activity of selected indole alkaloids. *Molecules*. 2013;18(6):6281-97.
44. Ayuko TA, Njau RN, Cornelius W, Leah N, Ndiege IO. In vitro antiplasmodial activity and toxicity assessment of plant extracts used in traditional malaria therapy in the Lake Victoria Region. *Memórias do Instituto Oswaldo Cruz*. 2009;104(5):689-94.
45. Schwikkard S, van Heerden FR. Antimalarial activity of plant metabolites. *Natural Product Reports*. 2002;19(6):675-92.
46. Muthaura C, Keriko J, Mutai C, Yenesew A, Gathirwa J, Irungu B, et al. Antiplasmodial potential of traditional phytotherapy of some remedies used in treatment of malaria in Meru–Tharaka Nithi County of Kenya. *Journal of ethnopharmacology*. 2015;175:315-23.
47. Mpiana P, Ngbolua K, Mudogo V, Tshibangu D, Atibu E, Mbala B, et al. The potential effectiveness of medicinal plants used for the treatment of Sick cell Disease in the Democratic Republic of Congo folk medicine: A review. *Progress in Traditional and Folk herbal medicine*. 2012;1:1-11.
48. Stangeland T, Alele PE, Katuura E, Lye KA. Plants used to treat malaria in Nyakayojo sub-county, western Uganda. *Journal of ethnopharmacology*. 2011;137(1):154-66.
49. Didier DS, Emmanuel MM, Alfred N, France KM, Lagarde BJ. Ethnobotanique et phytomédecine des plantes médicinales de Douala, Cameroun. *Journal of Applied Biosciences*. 2011;37:2496-507.
50. Anywar G, van't Klooster CI, Byamukama R, Wilcox M, Nalumansi PA, de Jong J, et al. Medicinal plants used in the treatment and prevention of malaria in Cegere Sub-County, Northern Uganda. *Ethnobotany Research and applications*. 2016;14:505-16.
51. Petersen I, Gabryszewski SJ, Johnston GL, Dhingra SK, Ecker A, Lewis RE, et al. Balancing drug resistance and growth rates via compensatory mutations in the *P. falciparum* chloroquine resistance transporter. *Molecular microbiology*. 2015;97(2):381-95.
52. Dhingra SK, Redhi D, Combrinck JM, Yeo T, Okombo J, Henrich PP, et al. A variant PfCRT isoform can contribute to *Plasmodium falciparum* resistance to the first-line partner drug piperazine. *MBio*. 2017;8(3):e00303-17.
53. Blasco B, Leroy D, Fidock DA. Antimalarial drug resistance: linking *Plasmodium falciparum* parasite biology to the clinic. *Nature medicine*. 2017;23(8):917.
54. Costa GL, Amaral LC, Fontes CJF, Carvalho LH, de Brito CFA, de Sousa TN. Assessment of copy number variation in genes related to drug resistance in *Plasmodium vivax* and *Plasmodium falciparum* isolates from the Brazilian Amazon and a systematic review of the literature. *Malaria journal*. 2017;16(1):152.
55. Tshibangu PT, Kapepula PM, Kapinga MK, Mukuta AT, Kalenda DT, Tchinda AT, et al. Antiplasmodial activity of *Heinsia crinita* (Rubiaceae) and identification of new iridoids. *Journal of ethnopharmacology*. 2017;196:261-6.
56. Kiplagat DM, Akala HM, Liyala PO, Wangui JM, Odhiambo RA, Omolo JO. Antiplasmodial activity of flavan derivatives from rootbark of *Cassia abbreviata* Oliv. *Journal of Saudi Chemical Society*. 2016;20:S140-S4.

57. Appiah-Opong R, Nyarko A, Dodoo D, Gyang F, Koram K, Ayisi N. Antiplasmodial activity of extracts of *Tridax procumbens* and *Phyllanthus amarus* in in vitro *Plasmodium falciparum* culture systems. *Ghana medical journal*. 2011;45(4).
58. Owuor B, Ochanda J, Kokwaro J, Cheruiyot A, Yeda R, Okudo C, et al. In vitro antiplasmodial activity of selected Luo and Kuria medicinal plants. *Journal of ethnopharmacology*. 2012;144(3):779-81.
59. Okokon JE, Antia BS, Mohanakrishnan D, Sahal D. Antimalarial and antiplasmodial activity of husk extract and fractions of *Zea mays*. *Pharmaceutical biology*. 2017;55(1):1394-400.
60. Zemicheal G, Mekonnen Y. Antiplasmodial activity of *Vernonia adoensis* aqueous, methanol and chloroform leaf extracts against chloroquine sensitive strain of *Plasmodium berghei* in vivo in mice. *BMC research notes*. 2018;11(1):736.
61. Sangian H, Faramarzi H, Yazdinezhad A, Mousavi SJ, Zamani Z, Noubarani M, et al. Antiplasmodial activity of ethanolic extracts of some selected medicinal plants from the northwest of Iran. *Parasitology research*. 2013;112(11):3697-701.
62. Soma A, Sanon S, Gansané A, Ouattara LP, Ouédraogo N, Nikiema J-B, et al. Antiplasmodial activity of *Vernonia cinerea* Less (Asteraceae), a plant used in traditional medicine in Burkina Faso to treat malaria. *African Journal of Pharmacy and Pharmacology*. 2017;11(5):87-93.